

تأثیر فلزات سنگین روی و کادمیم بر رشد و برخی صفات فیزیولوژیک

اسفناج

The impact of Zn and Cd heavy metals on the growth and some physiological characteristics of spinach

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چکیده

غلظت بالای فلزات سنگین باعث ایجاد اختلال در فرایندهای متابولیکی رشد گیاهان شده و حتی باعث مرگ آن‌ها می‌شود. گیاهان آستانه تحمل متفاوتی به فلزات سنگین نشان می‌دهند به همین منظور تأثیر غلظت‌های مختلف فلزات سنگین روی و کادمیم (صفر، ۲ و ۴ میلی گرم در لیتر) در دو رقم اسفناج (بذر معمولی و خاردار) بر روی برخی صفات مورفولوژیکی، فیزیولوژیکی و محتوای عناصر، در قالب آزمایش فاکتوریل بر مبنای طرح کاملاً تصادفی با سه تکرار مورد بررسی قرار گرفت. نتایج نشان داد برخی صفات مورفولوژیکی گیاه فقط تحت تأثیر مستقل فلزات و نوع رقم قرار دارند به طوری که کمترین وزن تر برگ و ارتفاع گیاه در تیمار چهار میلی گرم در لیتر کادمیم و بیشترین مقدار این دو صفت در تیمار روی دو میلی گرم در لیتر مشاهده شد. بالاترین وزن خشک گیاه در رقم خاردار و بیشترین ارتفاع گیاه در رقم بذر معمولی ثبت گردید. اثرات متقابل نوع رقم و فلزات سنگین در برخی صفات فیزیولوژیکی مانند محتوای مواد جامد محلول، محتوای کاتالاز و سوپراکسید دیسموتاز معنی‌دار بود. بیشترین فعالیت آنزیم کاتالاز در تیمار ۲ میلی گرم در لیتر کادمیم در بذر معمولی و بیشترین میزان آنزیم سوپراکسید دیسموتاز در تیمار ۴ میلی گرم در لیتر روی و ۲ میلی گرم کادمیم در بذر معمولی مشاهده شد. فلز روی تا دو میلی گرم در لیتر باعث افزایش رشد و نمو اسفناج گردید ولی بیشتر از این مقدار مانند فلز کادمیم باعث کاهش عملکرد و افزایش مقدار آنزیمهای آنتی اکسیدانی شد. تمامی محتوای عناصر اندازه گیری شده اثر متقابل معنی‌داری نشان دادند. با افزایش کادمیم جذب کلسیم، پتاسیم، روی، منگنز و منیزیم توسط گیاه کاهش یافت به طوری که کمترین مقدار تمامی عناصر اندازه گیری شده در چهار میلی گرم در لیتر کادمیم در بذر خاردار مشاهده گردید. نتایج بدست آمده از تعیین غلظت زیستی نشان داد که محتوای کادمیم برگ اسفناج خاردار و معمولی بین ۱۴۵ تا ۱۶۷ میلی گرم در کیلوگرم متغیر بود و اسفناج می‌تواند نماینده خوبی برای استفاده در زیست پالایی فلز کادمیم باشد.

واژه‌های کلیدی

اسفناج،
صفات فیزیولوژیک،
فلز سنگین،
آنزیم

Introduction

With the development of industrial zones and the huge population growth, many parts of the world especially industrial regions and the places with heavy mining industry and even the areas with prolonged agricultural patterns are facing progressive pollution problems (Liu *et al.* 2010). The concentrations of heavy metals higher than normal threshold in the soils gradually restrict the plant growth, food production and microbial populations in the soil. Heavy metals have direct deteriorative effects on ecosystems. Variations in the biochemical, physiological and metabolism of plants under stressful conditions are frequently reported. Some heavy metals, like Co, Cr, Fe, Mn, Ni, Cu, Cd and Zn are un-substitutable components of pigments or enzymes in plants and vegetation. So, they are essential elements in very trace amounts. (Srivastava *et al.* 2017; Ali *et al.* 2015). The presence of high dosages of Zn and Cd interfere soil organisms at the genes level and checks the enzymes and microbial activity in the soil (Liu *et al.* 2010). Furthermore, heavy metals hinder the metabolic pathways even at the cellular level. Zn-toxicity drastically affects dehydrogenase, urease, and β - glycosidase activity. Meanwhile, entering Zn^{2+} in the human food chains hurts pancreas and disturb proteins metabolism (Zhi-Xin *et al.* 2006). Whereas, Cd toxic levels prevents the activity of alkaline-phosphatase at the soil environment (Zhi-Xin *et al.* 2006). The plant type is the dominant criteria depicting tolerance to heavy metal contamination. Plants intrinsically have several mechanisms to combat and tolerate the heavy metals toxicity; prevention of the minerals entrance into the cells by their attachment to the cell wall, and producing low molecular weight enzymatic antioxidants (Shu *et al.* 2011) superoxide and H_2O_2 (Amaraathared *et al.* 2011). A research conducted on spinach revealed that under Cd high availability, the element accumulation was at the toxic levels in the roots and shoots of plants (Eisazadeh Lazarjan *et al.* 2016). However, with higher-growth rate and higher yields, spinach could be a good phytoremediation candidate for the Cd polluted soils (Eisazadeh Lazarjan *et al.* 2015). Cd influence the growth and physiological response of radish by the impact on growth characteristics and finally yield in main part by the negative effects on proline, anthocyanin and phenolic content (Moridian Pidosti *et al.* 2018). In *Brassica napus*, Cd inhibited the seed germination by th structural defects caused on seed cells (Ali *et al.* 2015). Moreover, in spinach, the high Zn, Cd, Ni and Cu concentration recorded the growth potential via the antioxidant enzymes deterioration and the toxic

effects they have been on the activated CAT and the great increase in the SOD activity (Pandy *et al.* 2009) So, the identification and characterization of plants with the reasonable tolerance for the risky environments is greatly crucial. Compared to physico-chemical amendment procedures, phytoremediation is economic environment friend method to get-ride of heavy metals toxicity in the soil. The aim of the present study was to investigate the effects of Cd and Zn availability in the growing media on the growth of spinach (as a candidate for phytoremediation) in hope to have a detailed idea on the recommendation of this crop for the remedy of Zn- Cd contaminated soils.

MATERIALS AND METHODS

The experiment was conducted during the year 2017 at the Research Greenhouse of Azarbaijan Shahid Madani University, Iran. The light intensity was $400 \mu\text{mol m}^{-2}\text{s}^{-1}$ under a 16: 8, day: night regime, and $20\text{-}25^{\circ}\text{C}$ day/night temperature. Two seed types (common and thorny) were used. The seeds were surface sterilized by sodium hypochlorite 10 % for 30 min and were planted at 5 liter pots filled with medium sized perlite. The plants were nourished by the Hoagland's nutrient solution (Hassanpouraghdam *et al.* 2019). pH of the nutrient solution was adjusted at 5.6. To avoid the salinity setup, the growing media were weekly washed with tap water. After the establishment of plants, heavy metals (Zn and Cd) treatments (0, 2 and 4 mg L^{-1}) were imposed on plants. The plants were sampled for analysis before flowering.

Plant fresh and dry weight: The fresh and dry weight of aerial parts were recorded by a digitl scale (BB141, Boeco, Germany).

Minerals content: Minerals content was quantified by the method of Honarjoo *et al.* (2013).

Enzymes activity: H_2O_2 content was traced by Amaraathared *et al.* (2011) and superoxide dismutase was evaluated by Minami and Yoshilcawa (1997) based on the assay of light dependent nitro- blue-tetra Zolium reduction at 560 nm as well as catalase by the method of Luhova *et al.* (2003).

The assessment of Bio-Concentration Factor (BCF) in the roots and leaves: The trait was assayed as: the concentration added to the growing medium (mgkg^{-1}) / the mean elemental concentration at plant tissue (mg kg^{-1}) (Ghosh and Sing, 2005).

Data analysis: The data were analyzed by MSTATC software. Means were compared by Duncan's multiple range test.

RESULTS AND DISCUSSION

Plant fresh and dry weight

The leaves fresh weight was affected by the heavy metals type (Table 1). The highest dry weight of plant was belonged to 2 mg L⁻¹ of Zn and the least plant fresh weight was recorded at 4 mg L⁻¹ Cd (Table 3). Thorny spinach type attained the highest plant fresh weight (Table 2). Photosynthesis reaction is more sensitive to Cd toxicity and the Cd high-availability greatly reduces the stomatal conductance, chlorophyll content, disturbs photosynthetic enzymes activity, and interferes electron transport in photosystems I and II. It seems that the efficiency of some plants in Cd accumulation is dependent upon the cells detoxification (Nikolic *et al.* 2014). Ghasemi and Shahabi (2011) reported that Cd availability reduced the growth and yield of tomato. The main idea about is lessening cell expansion and/or cell division prevention. Furthermore, Cd interferes photosynthesis and respiration and, by impairing CO₂ influx or by impacting the enzymes of carbon reducing cycle in photosynthetic apparatus reduces the photosynthetic rate and dry matter production (Penge *et al.* 2008). Aravind and Prasad (2004) reported that Cd reduced chlorophyll biosynthesis by the impact on δ -aminolevulinic acid dehydratase. Besides, Cd reduces protochlorophyll reductase by its effects on sulphhydryl radicles and by reducing the 5-amino- luvulinic acid (ALA) levels. This later acid is a precursor for the formation of tetrapyrrol in chlorophyll structure (Aravind and Prasad, 2004).

Plant height

Table 2 shows that the top height was recorded with common spinach seeds showing 22 % increase compared to thorny seed type. Heavy metals influenced plants height as well, and, the highest data was recorded in 2 and 4 mg L⁻¹ of Zn (Table 3). It seems that Zn concentration employed had no negative effects on plant height. Contrarily, in a study conducted on maize, the scientists reported that Zn application improved plant height, aerial parts dry weight, the relative water content and Zn and K content in aerial parts (Karmollachaab and Gharineh, 2013). Zn is a dominant micronutrient having pivotal roles in DNA, RNA, auxin and protein biosynthesis; its deficiency disturbs metabolism and deteriorates macromolecules like cell membranene achored proteins, chlorophylls, nucleic acids and enzymes of

IAA biosynthesis pathway and eventually, reduces growth potential and productivity of plants (Cakmak 2000, Broadley *et al.* 2007). Previous studies have verified that Cd modifies the meristematic cell division domains. The low Cd (1-4 μ mol) increases cell division and plant growth. However, with the higher concentrations, retard cell division, plant growth and genes expression pattern especially cyclin B1 genes are interfered; the gene which are actively involved in cell division behaviors (Liu *et al.* 2010).

H₂O₂ content

Hydrogen peroxide content was influenced by the interaction effects of cultivar and heavy metal treatment. The highest H₂O₂ content was recorded under 2 mg L⁻¹ Cd in common seeds (Table 4). Our results are the same as the reports of Vojodi Mehrabani *et al.* (2017) and Valizadeh *et al.* (2017). Cd stimulates H₂O₂ and subsequently hydroxyl radicals biogenesis and accelerates cell membrane lipids decomposition and peroxidation and hence, speeds up membrane deterioration and cell death (Hsu band Lao 2007). The toxic heavy metals availability stimulates the production of ROS molecules such as; superoxide, hydroxyl and hydrogen peroxide. These destructive molecules invade all types of vital macromolecules and demolish cell membrane leading to membrane breakdown and a very increased membrane leakage (Xiao *et al.* 2008).

Catalase activity

Catalase activity was significantly influenced by the interaction of cultivar and heavy metals level. The most recorded data was in 4 mg L⁻¹ Zn with thorny seed (Table 4). Vojodi Mehrabani *et al.* (2017) reported that, with Cd application, catalase and ascorbate peroxidase activities were increased in garden cress. They have reported that the higher SOD activity lead to higher H₂O₂ release and to keep the redox potential, H₂O₂ was decomposed into H₂O and oxygen by the action of catalase and peroxidase. Catalase has an important role in scavenging H₂O₂ into H₂O and O₂. Unyayar *et al.* (2005) reported that even H₂O₂ in high doses is toxic for the cells. However, with low concentration plays as a messenger in gene expression process related to tolerance. Catalase with overcoming on oxidative stress and by scavenging H₂O₂ prevents ROS molecules generation and so prevents protein and nucleic acids deterioration (Unyayar *et al.* 2005).

Table 1- ANOVA for the effects of cultivar and different levels of Zn and Cd on enzymic activity, H₂O₂ content, elemental content and morphological traits of *Spinacia oleracea* L.

Source of variation	de	K content	Ca content	Mg content	Mn content	Cd content	Zn content	Plant height	Arial part dry weight (g)	Arial part fresh weight (g)	Catalase activity	H ₂ O ₂ content	Superoxide dismutase activity
Seed type	1	10 ^{7**} ×40	10 ^{**} ×53	**39463	**1393	**61027	**18820	104*	0.002 ^{ns}	0.008*	17361 ^{ns}	0.005 ^{ns}	0.01 ^{ns}
Heavy metal treatment	4	10 ^{6**} ×33	10 ^{6**} ×22	724 ^{ns}	**265	**80195	**87433	30*	0.03*	0.002 ^{ns}	88302 ^{ns}	0.007*	0.05 ^{ns}
Seed type × Heavy metal treatment	4	10 ^{6**} ×13	10 ^{5**} ×51	**1747	**121	**14606	**4666	2.3 ^{ns}	0.004 ^{ns}	0.001 ^{ns}	164009*	0.012 ^{**}	0.11*
Error	18	2611729.19	10 ³ ×16	631	20	176.3	149	7.4	0.0001	0.001	54928	0.002	0.02
c.v(%)		6.3	1.1	1.01	6.8	9.3	2.8	14	6.8	7.9	12.9	6.6	5.9

ns,* and ** show non-significant and significant at P≤0.05 and P≤0.01, respectively.

Table 2- Mean comparison for the effects of cultivar on plant height and fresh weight of *Spinacia oleracea*.

Cultivars	Plant height (cm)	Arial part fresh weight (g)
Thorny seeds	13 ^b	22.4 ^a
Common seed	16.7 ^a	11.3 ^b

Similar letters in the columns are non-significant based on Duncan's test

Table 3- Mean comparison for the effects of heavy metal type on plant height and arial part fresh weight of *Spinacia oleracea*

Metal type	Plant height (Cm)	Arial part dry weight (g)
Control	11 ^b	0.46 ^b
2 mg L ⁻¹ Zn	16 ^a	0.56 ^a
4 mg L ⁻¹ Zn	16 ^a	0.40 ^b
2 mg L ⁻¹ Cd	11 ^b	0.44 ^b
4 mg L ⁻¹ Cd	9.3 ^c	0.33 ^c

Similar letters in the columns are non-significant based on Duncan's test.

Table 4- Mean comparison for the interaction effects of cultivar and different levels of heavy metals on enzyme activity and H₂O₂ content of *Spinacia oleracea* L.

Cultivar	Heavy metal treatment (mg L ⁻¹)	H ₂ O ₂ content (absorption changes min ⁻¹ mg ⁻¹ protein)	Superoxide dismutase activity (enzyme unit 1 mg ⁻¹ protein)	Catalase activity (μmol g ⁻¹ FWt)
Thorny seed	Control	0.06 ^b	0.15 ^{ab}	371 ^b
Thorny seed	2 mg L ⁻¹ Zn	0.02 ^b	0.10 ^b	343 ^b
Thorny seed	4 mg L ⁻¹ Zn	0.03 ^b	0.14 ^b	855 ^a
Thorny seed	2 mg L ⁻¹ Cd	0.06 ^b	0.22 ^b	189 ^b
Thorny seed	4 mg L ⁻¹ Cd	0.07 ^b	0.26 ^b	271 ^b
Common seed	Control	0.02 ^b	0.08 ^b	369 ^b
Common seed	2 mg L ⁻¹ Zn	0.04 ^b	0.17 ^b	314 ^b
Common seed	4 mg L ⁻¹ Zn	0.3 ^b	0.65 ^a	302 ^b
Common seed	2 mg L ⁻¹ Cd	0.21 ^a	0.36 ^a	535 ^{ab}
Common seed	4 mg L ⁻¹ Cd	0.03 ^b	0.12 ^b	238 ^b

Similar letters in the columns are non-significant based on Duncan's test.

SOD activity

The highest data for SOD activity was recorded in common seed spinach with 2 and 4 mg L⁻¹ of Zn and thorny seed × control plants. In thorny seed cultivars, there was no difference in SOD activity considering the studied traits (Table 4). SOD is a metalloenzyme having Fe, Cu, Zn or Mn in its prosthetic active site under higher Cd availability, Fe, Zn and Mn content in plant tissue was declined and seems that SOD activity declined reduction in Cd exposed plants is a response to the deficiency of mineral elements which are essential for the activity of the related enzymes (Chamseddine *et al.* 2009). The Same data have been reported by Pandey *et al.* (2009) in spinach. In a research conducted by Sobkowiak and Jaunna (2003), Cd was replaced by Zn. Zinc is a pivotal element for binding the transcription factor in regulating sites of several genes. It seems that higher Cd concentration goes to the reduced transcription factors availability and hence, the declined transcriptomes of SOD and APX. Moreover, Cd modifies the DNA methylation pattern, diminishes RNA biosynthesis or prevents

RNA polymerase action in the transcription of the related genes (Pierron *et al.* 2014).

Cd content

The highest Cd content was belonged to thorny seeds × leaf sample of 4 mg L⁻¹ of Cd as well as in common seed × leaf sample of 4 mg L⁻¹ of Cd (Table 5). In the study of Vojodi Mehrabani *et al.* (2017) with more Cd availability, its amounts were higher in the plants aerial parts. Cd by passing the apoplastic route or by the simplistic pathway enters the root cells and by the xylem sap transfers to the aerial plant parts (Nikolic *et al.* 2014). Cd negatively impacts cell division and enlargement, plant growth and productivity. Furthermore, Cd prevents rubisco enzyme activity; the key enzyme in Calvin cycle, and consequently retards transpiration, minerals uptake and distribution and, N and P metabolism (Manna, 2013). In the sensitive plants, Cd alters the membrane permeability and retards or prevents minerals intake and impedes appropriate concentration of minerals inside cells. Genetic variability in Cd accumulation in plants is related to the physiological aspects of plants such as; the diverse ability of plants roots in Cd

absorption, diversity in the translocation of Cd and the different ability in Cd accumulation.

Zn content

The highest Zn content was recorded for thorny seed × leaf samples under 4 mg L⁻¹ Zn and in common seeds × 2 mg L⁻¹ Zn (Table 5). The same results on the reduction of Zn absorption in plants in response to Cd availability have been reported by Yildiz (2005). Alio *et al.* (2015) reported that with increasing heavy metals (Zn and Cd) in plant growing medium the absorption of plant nutrition was decreased in spinach. Furthermore, Pandey *et al.* (2009) noted that the long time exposure to heavy metals of Zn, Cd and Ni retarded the growth of plants and led to chlorosis as well as reduced the ascorbate content of plants. Zn at low concentration is an essential micro-nutrient, but at higher concentration reduce plant growth, induces chlorosis and alters the chlorophylls structure (Ali *et al.* 2015). Moreover, Chaoui *et al.* (1997) noted that, with any Zn availability, protein and some antioxidant enzymes biosynthesis was improved. These protein molecules bind to metal elements and by generating metallothionein (protein metal complexes) neutralize the toxic effects of elements.

Potassium content

The highest K content was documented in thorny seed × leaf sample of 4 mg L⁻¹ Zn, root sample of thorny seed and common spinach root sample × 2 and 4 mg L⁻¹ Zn (Table 5). Cd has high affinity with sulfhydryl, hydroxyl and nitrogen containing ligands and by this way inactivates several enzymes. Furthermore, Cd interferes photosynthesis, respiration and other metabolic processes. The major reason in Cd toxicity is its interaction with essential mineral elements which hinders the balanced uptake and distribution of essential elements and goes to vital elements deficiency (Ciecko 2004). Cd availability reduced the uptake of Ca²⁺, Mg²⁺ and K⁺ in plant (Vojodi Mehrabani *et al.* 2017; Valizadeh *et al.* 2017).

Calcium content

Thorny seed × leaf sample of control plants hold the highest Ca²⁺ content (Table 5). The diminished

Ca²⁺ uptake in response to Cd availability has been reported by Gussarsson *et al.* (1996) Growth retardation is the first response to the Cd toxicity. Ghasemi and Shahabi (2011) reported that in maize plants, with any Cd concentration increase, the absorption of N, P, K, Zn Cu and Na declined. Similar result about decreasing in elemental content was reported by Alio *et al.* (2015) in spinach. Increasing Ca²⁺ concentration inside plant is an efficient mechanism against Cd toxicity (Ghasemi and Shahabi, 2011).

Mg content

Mg²⁺ had the highest concentration in thorny seeds × leaf sample of control plants and in common seeds × leaf sample of 2 mg L⁻¹ Zn treatments (Table 5). Cd toxicity and higher availability declined the absorption of Mn, Fe and Cu in tomato leaves (Ghasemi and Shahabi, 2011). Cd is involved in the movement and translocation of micro nutrients to the leaves. It seems that the major reason for nutrients concentration loss in the Cd rich environment is because of the competition for the binding sites or the differential selectivity of elemental carriers toward the cells. The differences in the potential of Cd uptake by the roots and the subsequent translocation and accumulation in the aerial parts is a pivotal factor in describing the differences between genotypes in their tolerance to the heavy metals. In the research conducted by Nikolic *et al.* (2014), the sufficient mg²⁺ content in plant tissues was essential to compensate the toxic Cd effects.

Mn²⁺ content

The highest Mn²⁺ concentration was recorded in common seed × leaf samples of 2 and 4 mg L⁻¹ of Zn (Table 5). Cd availability negatively impacted Mn²⁺ uptake. Rubisco activity is thoroughly sensitive to Cd toxicity. Cd in the photolysis reaction, substitutes Mn²⁺ ion and hinders the water complex breaking (Stobar *et al.* 1985). Mn²⁺ uptake improvement in Marvdasht wheat under 4 mg L⁻¹ of Cd could be described to the lower concentration of Cu²⁺, Zn²⁺ and Fe²⁺. Cd competes with Mn²⁺ in protein carriers of cell membranes (Penge *et al.* 2008).

Table 5- Mean comparison for the interaction effects of cultivar and different levels of heavy metal on elemental content (mg Kg⁻¹) of *Spinacia oleracea* L.

Cultivar	Heavy metal treatment	Ca content	K content	Zn content	Mn content	Mg content	Cd content
Thorny seeds	Control (leaf)	19.4 ^a	23.8 ^{b-d}	0.30 ^{hi}	0.06 ^{d-g}	2.5 ^a	-
Thorny seeds	2 mg L ⁻¹ cd in leaf sample	12.5 ^f	22.1 ^{b-d}	0.44 ^{fg}	0.06 ^{d-f}	2.4 ^{c-f}	0.33 ^b
Thorny seeds	4 mg L ⁻¹ cd in leaf sample	15.6 ^d	18.0 ^f	0.26 ^{hi}	0.06 ^{b-d}	2.5 ^{b-e}	0.37 ^a
Thorny seeds	2 mg L ⁻¹ Zn in leaf sample	15.3 ^e	21.5 ^{b-f}	0.54 ^b	0.06 ^{b-d}	2.5 ^{b-d}	-
Thorny seeds	4 mg L ⁻¹ Zn in leaf sample	5.5 ⁱ	33.0 ^a	0.59 ^a	0.05 ^{hi}	2.4 ^{c-f}	-
Thorny seeds	Control (root)	4.8 ^k	25.0 ^b	0.30 ^{hi}	0.06 ^{e-h}	2.4 ^{d-g}	-
Thorny seeds	2 mg L ⁻¹ cd in root sample	4.82 ^k	25.0 ^b	0.30 ^{hi}	0.06 ^{e-h}	2.4 ^g	0.09 ^e
Thorny seeds	4 mg L ⁻¹ cd in root sample	6.11 ^h	17.7 ^f	0.47 ^{de}	0.05 ^{hi}	2.4 ^{d-g}	0.12 ^d
Thorny seeds	2 mg L ⁻¹ zn in root sample	5.15 ^j	30.0 ^a	0.46 ^{d-f}	0.05 ^{g-i}	2.4 ^{d-g}	-
Thorny seeds	4 mg L ⁻¹ zn in root sample	6.25 ^{gh}	20.0 ^{b-e}	0.44 ^{fg}	0.06 ^{d-g}	2.4 ^{d-g}	-
Common seed	Control (leaf)	18.96 ^b	21.0 ^{c-f}	0.29 ⁱ	0.06 ^{c-e}	2.3 ^{f-h}	-
Common seed	2 mg L ⁻¹ cd in leaf sample	12.5 ^f	22.0 ^{b-f}	0.41 ^g	0.06 ^{e-g}	2.4 ^{fg}	0.29 ^c
Common seed	4 mg L ⁻¹ cd in leaf sample	17.03 ^c	20.0 ^{d-f}	0.48 ^{de}	0.05 ^{f-i}	2.3 ^{f-h}	0.38 ^a
Common seed	2 mgL ⁻¹ Zn in leaf sample	15.64 ^d	18.1 ^{ef}	0.59 ^a	0.08 ^a	2.5 ^{ab}	-
Common seed	4 mg L ⁻¹ Zn in leaf sample	15.50 ^{de}	21.0 ^{c-f}	0.51 ^c	0.07 ^{ab}	2.5 ^{b-d}	-
Common seed	Control (root)	5.44 ⁱ	22.8 ^{b-e}	0.32 ^h	0.05 ^{f-i}	2.4 ^{fg}	-
Common seed	2 mg L ⁻¹ cd in root sample	4.95 ^{jk}	24.9 ^{bc}	0.32 ^h	0.06 ^{e-h}	2.4 ^{fg}	0.09 ^e
Common seed	4 mg L ⁻¹ cd in root sample	6.14 ^{gh}	21.8 ^{b-e}	0.48 ^{cd}	0.04 ⁱ	2.4 ^{fg}	0.12 ^d
Common seed	2 mg L ⁻¹ zn in root sample	5.13 ^j	29.0 ^a	0.45 ^{ef}	e-i 0.05	2.4 ^{e-g}	-
Common seed	4 mg L ⁻¹ zn in root sample	6.43 ^g	29.9 ^a	0.41 ^g	0.06 ^{b-e}	2.4 ^{d-g}	-

Similar letters in the columns are non-significant based on Duncan's test.

Bio-concentration factor

Bio-concentration factor is the potential of plants in accumulation of a special metal element from the containing medium. Variations in bio-concentration factor is dependent upon the bio-accumulation per biomass of a given plant as well as on the element concentration. $BFC \geq 1$ means that, the plant is adequately potential in the absorption and

translocation of element (Ghosh and Singh, 2005). The plants species have different behaviors in response to heavy metals. Based on the results obtained (Table 6), Cd content of thorny and common spinach leaves was between $145-167 \text{ mg kg}^{-1}$. The least Zn content was recorded in 4 mg L^{-1} of Cd in thorny seed spinach leaves. The results of the present experiment are in line with the findings of Vojodi Mehrabani *et al.* (2016).

Table 6- The means for Bio-Concentration factor (BCF) of Zn and Cd in *Spinacia oleracea* L.

Cultivar	Heavy metal treatment	Zn content (g Kg^{-1})	Cd content (g Kg^{-1})
Thorny seeds	Control (leaf)	0.30	-
Thorny seeds	2 mg L^{-1} cd in leaf sample	0.22	0.16
Thorny seeds	4 mg L^{-1} cd in leaf sample	0.14	0.09
Thorny seeds	2 mg L^{-1} Zn in leaf sample	0.27	-
Thorny seeds	4 mg L^{-1} Zn in leaf sample	0.08	-
Thorny seeds	Control (root)	0.30	-
Thorny seeds	2 mg L^{-1} cd in root sample	0.15	0.04
Thorny seeds	4 mg L^{-1} cd in root sample	0.11	0.03
Thorny seeds	2 mg L^{-1} zn in root sample	0.23	-
Thorny seeds	4 mg L^{-1} zn in root sample	0.11	-
Smooth seed	Control (leaf)	0.29	-
Smooth seed	2 mg L^{-1} cd in leaf sample	0.20	0.14
Smooth seed	4 mg L^{-1} cd in leaf sample	0.12	0.09
Smooth seed	2 mg L^{-1} Zn in leaf sample	0.28	-
Smooth seed	4 mg L^{-1} Zn in leaf sample	0.12	-
Smooth seed	Control (root)	0.32	-
Smooth seed	2 mg L^{-1} cd in root sample	0.16	0.04
Smooth seed	4 mg L^{-1} cd in root sample	0.12	0.03
Smooth seed	2 mg L^{-1} zn in root sample	0.22	-
Smooth seed	4 mg L^{-1} zn in root sample	0.10	-

Similar letters in the columns are non-significant based on Duncan's test.

Conclusions

Accumulation rate of heavy metals in plants is quite dependent on species and growing conditions. The overall results indicated the positive effects of Zn on dry weight, plant height, SOD content, as well as on Zn^{2+} and K^{+} content. Cd toxicity stress increased

CAT and H_2O_2 content of plants. Overall, for growing spinach as a vegetable, Zn application would be a promising way to improve the growth and yield of plant. Otherwise, for phyto-remediation of Cd polluted areas, spinach would be again a promising candidate owing to the fast-growing habit and multi harvest nature of the plant during a year.

REFERENCES

- Ali B, Deng X, Hu X, Gill RA, Ali S, Wang S, Zhou W. 2015. Deteriorative effects of cadmium stress on antioxidant system and cellular structure in germinating seeds of *Brassica napus* L. Journal of Agricultural Science and Technology 17: 63-74.
- Alio N, Sardar K, Said M, Salma Kh, Sadia A, Sadaf S, Toqeer A, Miklas S. 2015. Toxicity and Bioaccumulation of heavy metals in spinach grown in a controlled environment. International Journal of Environmental Research and Public Health 12(7): 7400-7416.
- Amaraathared Y, Lokesh U, Venkatesh B, Sudhakar C. 2015. Pb-stress induced oxidative stress caused alterations in antioxidant efficacy in two groundnuts (*Arachis hypogaea* L.) cultivars. Agricultural Science 6(10): 1283-1297.
- Aravind P, Prasad NBC. 2004. Zink protects chloroplasts and associated photochemical function in cadmium exposed *Ceratophyllum demersum* L. a fresh water macrophage. Plant Science 166 (5): 1231-1327.
- Broadley MY, Philip JW, Hammond JP, Zelko I, Alexander L. 2007. Zinc in plants. New Phytologist 173: 677-702.
- Cakmak I. 2000. Possible roles of zinc in protecting plant cell from damage by reactive oxygen species. The New phytologist 146: 185-205.
- Ciecko Z, Kalembasa S, Wyszowski M, Rollas E. 2004. Effect of soil contamination by cadmium on potassium uptake by plant. Polish Journal of Environment Studies 13. 333-337.
- Chamseddine M, Wided BA, Guy H, Marie-edith C, Fatma J. 2009. Cadmium and copper induction of oxidative stress and anti-oxidative response in tomato (*Solanum lycopersicon*) leaves. Plant Growth Regulation 57: 89-99.
- Chaoui A, Mazhoudi S, Ghor MH, Elferjani E. 1997. Cadmium and zinc induction if lipid peroxidation and effect on antioxidant enzyme activates in bean (*Phaseolus vulgaris* L.). Plant Science 127: 139-147.
- Eisazadeh Lazarjan S, Asadi Kapourchal S, Homae M. 2015. Phyto-extraction and estimating optimal time for remediation of Cd contaminated soils by spinach (*Spinacia oleracea* L.). Agroecology 6(4): 916-926. (In Farsi with English abstract).
- Ghasemi Z, Shahabi A. 2011. The effect of Cd on physiological traits, growth characteristics and minerals content of tomato. The Science and Technology of Greenhouse Crops 2: 55-65.
- Ghosh M, Singh SP. 2005. A review on phytoremediation of heavy metals and utilization of its byproducts. Asian Journal on Energy and Environment 6 (4): 214-231.
- Gussarsson M, Asp H, Adalsteinsson S, Jensen P. 1996. Enhancement of cadmium effects on growth and nutrient composition of birch (*Betula pendula*) by buthionine sulfoximine. Botany 47: 211-215.
- Hassanpouraghdam MB, Vojodi Mehrabani L, and Tzortzakakis N. 2019. Foliar application of nano-zinc and iron affects physiological attributes of *Rosmarinus officinalis* and quietens NaCl salinity depression. Journal of Soil Science and Plant Nutrition 16: 1-11.
- Honarjoo N, Hajrasuliha SH, Amini H. 2013. Three plants in absorption of ions from different natural saline and sodic soils. International Journal of Agriculture and Crop Science 6: 988-993.
- Hsu YT, Lao CH. 2007. Cadmium-induced oxidative damage in rice leave introduced by polyamines. Plant and Soil 291: 27-37.
- Karmollachaab A, Gharineh MH. 2013. Effect of zinc element on growth, yield components and some physiological characteristics of maize under NaCl salinity stress. Iranian Journal of Field Crops Research 11(3): 446-453.
- Liu J, Cao C, Wong M, Zhang Z, Chai Y. 2010. Variation between rice cultivars in iron and manganese plaque on root and relation with plant cadmium uptake. Environmental Science 22 (7): 1067-1072.
- Luhova L, Lebeda A, Hederorva D, Pec P. 2003. Activities of oxidase, peroxidase and catalase in seedlings of *Pisum sativum* L. under different light conditions. Plant Soil and Environment 49 (4): 151-157.

- Manna, D. 2013.** Growth, Yield and Bulb Quality of Onion in Response to Foliar Application of Boron and Zinc. SAARC Journal of Agriculture 11(1): 149-153.
- Minami M, Yoshil-Cawa H. 1997.** A simplified method of superoxide dismutase activity for clinical use. Clinica Chimica Acta 92: 337-42.
- Moridian Pirdost Kh, Movahedi Z, Rostami M, 2018.** Effects of cadmium stress on morpho-physiological traits in garden cress and radish in an aeroponic system. Iranian journal of plant physiology 9(1): 2591-2599.
- Nikolic N, Borisev M, Pajevic S, Zupunski M, Topic M, Arsenov D. 2014.** Responses of wheat and maize plants to cadmium toxicity in relation to magnesium nutrition. Acta Botanica Croatica 73(2): 359-373.
- Pandy N, Pathak GC, Pandey DK, Pandey R. 2009.** Heavy metals, Co, Ni, Cu, Zn and Cd produce oxidative damage and evoke differential antioxidant responses in spinach. Brazilian Society of Pant Phyyisology 21(2): 103-111.
- Penge K, Luo C, You W, Lian C, Shen Z. 2008.** Manges uptake and interaction with cadmium the hyperaccumulator. Phytolacca Americana 674-681.
- Pierron F, Baillon L, Sow M, Gotreau S, Gonzalez P. 2014.** Effect of low-dose cadmium exposure on DNA methylation in the endangered European eel. Environmental Science and Technology 48: 797-803.
- Sirvastava V, Sarkar A, Singh S, Singh P, de Araujo A.S.F, Singh RP. 2017.** Agroecological responses of heavy metal pollution with special emphasis on soil health and plant performances. Frontiers in Environmental Science 5:64-71.
- Shu X, Yin L, Zhang Q, Wang W. 2011.** Effect of Pb toxicity on leaf growth, antioxi d enzyme activities, and photosynthesis in cuttings and seedlings of *Jatropha curcas* L. Environmental Science and Pollution Research 19(3): 893-902.
- Sobkowiak R, Joanna D. 2003.** Cadmium-induced changes in growth and cell cycle gene expression in suspension-culture cells of soybean. Plant Physiology and Biochemistry 41: 767-772.
- Stobar A, Giffiths W, Ameen-Bukhari T, Sherwood R. 1985.** The effect of Cd on the biosynthesis of chlorophyll in leaves of barley. Physiologia Plantarum 63: 293-298.
- Unyayar S, Kele Y, Cekic FO. 2005.** The antioxidative response of two tomato species with different drought tolerances as a result of drought and cadmium stress combinations. Plant Soil and Environment 51(2): 57-64.
- Valizadeh Kamran R, Vojodi Mehrabani L, HassanpourAghdam MB. 2017.** Study of the effects of lead and cadmium on growth and enzymatic activity of *Spinacia oleracea* in in vitro condition. Genetic Engineering and Biosafety Journal 6(2): 257-268.
- Vojodi Mehrabani L, Valizadeh Kamran R, Fattahi N, Safardoost M. 2017.** The effects of lead on growth charactrastics and some physiological traits of gardencress (*Lepidium sativum*) under in vitro condition. Genetic Engineering and Biosafety Journal 6(1): 85-94.
- Xiao X, Tongbin C, Zhizhuang A, Mei L. 2008.** Potential of *Pteris vittata* L. for phytoremediation of sites co-contaminated with cadmium and arsenic: the tolerance and accumulation. Journal of Environmental Science 20:62-67.
- Yildiz N. 2005.** Response of tomato and corn plants to increasing Cd levels in nutrient culture. Pakistan Journal of Botany 37(3): 593-599.
- Zhi-Xin Y, Shu-Ging L, Da-Wei Z, Sheng-Dong F. 2006.** Effects of cadmium zinc and lead on soil enzyme activities. Journal of Environmental Science 18(6): 1135-1141.

The impact of Zn and Cd heavy metals on the growth and some physiological characteristics of spinach**LamiaVojodi Mehrabani^{*1}, RanaValizadeh Kamran² and Homa Mirzaei¹**

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Abstract

High concentrations of heavy metals disrupt the metabolic processes of plant growth and even cause plant death. Plants show different tolerance thresholds to heavy metals, therefore, the effect of different concentrations of heavy metals zinc (Zn) and cadmium (Cd) (zero, 2 and 4 mgL⁻¹) in two varieties of spinach (common seed and Thorny seeds) on some morphological, physiological traits and content of the elements were examined in the factorial experiment based on a completely randomized design with three replications. The results showed that some morphological traits of the plant are affected by metals and cultivar type independently, so that the lowest wet weight of leaves and plant height was observed in four mgL⁻¹ of Cd treatment and the highest amount of these two traits was observed in two mgL⁻¹ of Zn. The highest dry weight of the plant was recorded in the thorny cultivar and the highest plant height was recorded in the normal seed cultivar. The interaction of the type of cultivar and heavy metals was significant in some physiological traits such as soluble solids content, catalase content, and superoxide dismutase. The highest activity of catalase enzyme was observed in the treatment of 2 mgL⁻¹ of Cd in normal seeds and the highest amount of superoxide dismutase was observed in the treatment of 4 mgL⁻¹ of Zn and 2 mgL⁻¹ of Cd in normal seeds. Zinc metal up to 2 mgL⁻¹ increased the growth and development of spinach, but more than this amount, like cadmium metal, reduced yield and increased the amount of antioxidant enzymes. All content of the measured elements showed a significant interaction. As the cadmium increased, the plant's uptake of the Ca, K, Zn, Mn and Mg decreased, so that the lowest amount of all the elements measured in four mgL⁻¹ of Cd was observed in thorny seeds. The results of determining the biological concentration in this experiment showed that the cadmium content of thorny and common spinach leaves varied between 145 and 167 mgKg⁻¹ and spinach could be a good representative for use in cadmium phytoremediation.

Keywords: Enzyme activity, Heavy metal, Physiological characteristics, *Spinacia oleracea*